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# ***Elodea canadensis* under N and CO<sub>2</sub> Limitation: Adaptive Changes in Rubisco and PEPCase Activity in a Bicarbonate User**

By

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**K e y   w o r d s :** Aquatic plants, photosynthesis, fluorescence, bicarbonate assimilation,  
*Elodea canadensis*.

## S u m m a r y

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Diffusion of CO<sub>2</sub> in water is 10,000 times slower than in air. Because of this photosynthesis in submerged aquatic macrophytes is often limited by CO<sub>2</sub> availability. *Elodea canadensis* shows HCO<sub>3</sub><sup>-</sup> utilization under conditions of CO<sub>2</sub> limitation. A closely related species, *Hydrilla verticillata*, which also belongs to the Hydrocharitaceae, shows a C<sub>4</sub> like mechanism when grown under so-called summer conditions where low availability of CO<sub>2</sub> is expected. Here it is shown that a similar C<sub>4</sub>-like mechanism is possibly also present in *E. canadensis*. To study the possible interference between the two processes, *E. canadensis* was grown at low and high CO<sub>2</sub> and NO<sub>3</sub><sup>-</sup>, conditions expected to affect HCO<sub>3</sub><sup>-</sup> utilization. Moreover the effect was studied of these growth conditions on the occurrence of the C<sub>4</sub> state. A low G<sub>CO<sub>2</sub></sub> was observed in *E. canadensis* when it was grown under low CO<sub>2</sub> conditions indicative for plants in the C<sub>4</sub>-state. The low G<sub>CO<sub>2</sub></sub> was related to a change in the ratio Rubisco and PEPCarboxylase indicating a central role of these enzymes. The very same conditions induced HCO<sub>3</sub><sup>-</sup> utilization in this species. Both mechanisms thus operate in the same plant simultaneously. Growing *E. canadensis* at high NO<sub>3</sub><sup>-</sup>, 0.75 mM, had a strong stimulatory effect on O<sub>2</sub> production in the light both in low and high CO<sub>2</sub> grown plants, suggesting that it has no direct relation with HCO<sub>3</sub><sup>-</sup> utilization. The actual cause for this stimulatory effect of high N on photosynthesis is unclear.

## I n t r o d u c t i o n

Submerged aquatic macrophytes (SAM) are often faced with limitation of their photosynthesis by low availability of inorganic carbon, e.g. dissolved CO<sub>2</sub>.

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Diffusion of CO<sub>2</sub> in water is about 10,000 times slower than in air. Furthermore, the major component of the dissolved inorganic carbon (DIC) pool is HCO<sub>3</sub><sup>-</sup> at pH 7-9, common for natural waters. Since only CO<sub>2</sub> diffuses freely over the plasmamembrane to the cytoplasm, other inorganic carbon species have to be converted into CO<sub>2</sub> or need an active transport system across the plasmamembrane to contribute to carbon fixation. To describe the inorganic carbon system, the following terms are used: CO<sub>2</sub>, dissolved free CO<sub>2</sub> including H<sub>2</sub>CO<sub>3</sub>; HCO<sub>3</sub><sup>-</sup>, bicarbonate; CO<sub>3</sub><sup>2-</sup>, carbonate; DIC: sum of CO<sub>2</sub>, H<sub>2</sub>CO<sub>3</sub>, HCO<sub>3</sub><sup>-</sup> and CO<sub>3</sub><sup>2-</sup>.

The slow diffusion of gases in water also generates a high O<sub>2</sub> level in the photosynthesizing cells of submerged aquatic plants. The resulting low CO<sub>2</sub>/O<sub>2</sub> ratio near the Rubisco site is favorable to photorespiration and accordingly would lead to a low rate of carbon fixation. In the course of evolution several SAM responded to this challenge by development of mechanisms to use HCO<sub>3</sub><sup>-</sup> as an alternative carbon source and/or mechanisms to concentrate inorganic carbon (for review see BOWES & SALVUCCI 1989, PRINS & ELZENGA 1989). Mechanisms from the first category are associated with an increased potential for HCO<sub>3</sub><sup>-</sup> utilization either by active uptake of HCO<sub>3</sub><sup>-</sup> in the leaf cells or by conversion of HCO<sub>3</sub><sup>-</sup> into CO<sub>2</sub>. The second category are CO<sub>2</sub>-concentrating mechanisms (CCM), based on a C<sub>4</sub>- or CAM-like metabolism. Often these mechanisms of enriching the cell with CO<sub>2</sub> are accompanied by an increased carbonic anhydrase (CA) activity in the apoplast of leaf cells.

Models for HCO<sub>3</sub><sup>-</sup> utilization in fresh water macrophytes are generally based on active H<sup>+</sup> extrusion into the apoplast driven by a plasmalemma bound ATPase. Two mechanisms have been proposed. The first described mechanism is a light dependent acidification of the apoplast and unstirred layer near the leaf surface (PRINS & al. 1982). This acidification results in a shift of the equilibrium between HCO<sub>3</sub><sup>-</sup> and CO<sub>2</sub> in the direction of CO<sub>2</sub> which then diffuses into the leaf cells. To balance the loss of H<sup>+</sup> there is an H<sup>+</sup> influx or OH<sup>-</sup> efflux at another part of the leaf. There is no agreement if the alkaline zones result from H<sup>+</sup> influx or OH<sup>-</sup> efflux or by another process. For convenience the expression H<sup>+</sup> influx is used here.

This mechanism has now been observed in a number of fresh water angiosperms. E.g., in *Elodea*, *Egeria*, and *Potamogeton* there generally is an acidification at the abaxial side of the leaf to pH values of 4-5 and an alkalinization at the adaxial side to pH values of 9-11 (PRINS & ELZENGA 1989). A second mechanism has been proposed for a.o. the Characeae. The basis for this latter mechanism is HCO<sub>3</sub><sup>-</sup> transport by symport with H<sup>+</sup>, driven by the proton motive force and accompanied by acid regions of H<sup>+</sup> efflux and alkaline regions of H<sup>+</sup> influx (LUCAS 1985).

A C<sub>4</sub>-like metabolism has been proposed for *Hydrilla verticillata* and other SAM. Typical C<sub>4</sub>-characteristics are induced in *Hydrilla* when grown under conditions normally expected to lead to high rates of photorespiration. SAM showing this C<sub>4</sub>-like metabolism are, like their terrestrial counterparts, characterized by a low CO<sub>2</sub>-compensation point ( $J_{CO_2}$ ), a photosynthesis relatively insensitive to O<sub>2</sub> and a low Rubisco/PEPCase (phosphoenolpyruvate) activity ratio (R/P), due to a high PEPCase activity (SALVUCCI & BOWES 1981, HOLADAY & al. 1983, BOWES &

SALVUCCI 1989). In this species, Rubisco is localized in the chloroplast and PEP-Case in the cytoplasm (REISKIND & al. 1989). After fixation of inorganic carbon by PEPCase in the cytoplasm, the concentrating of DIC occurs most likely in the chloroplast (REISKIND & al. 1997).

In *Elodea canadensis* acidification is induced by conditions of high light and low DIC in plants previously grown under conditions favorable to photorespiration. Acidification is absent in *Elodea* plants grown under high CO<sub>2</sub> conditions (ELZENGA & PRINS 1989). This observation led to the question: is acidification in *Elodea* accompanied by C<sub>4</sub>-characteristics, such as a low  $\Gamma_{CO_2}$  and a low R/P, as observed in *Hydrilla*. A low  $\Gamma_{CO_2}$  could facilitate the diffusion of CO<sub>2</sub> into the leaf cells, following the conversion of HCO<sub>3</sub><sup>-</sup> into CO<sub>2</sub> by the process of acidification (PRINS & al. 1982).

It is generally thought that the main physiological effect of HCO<sub>3</sub><sup>-</sup> utilization is an increase of the rate of carbon fixation. It may also contribute to suppression of photorespiration through an increased availability of CO<sub>2</sub> and thereby, to a more efficient use of nitrogen (BEARDALL & al. 1982, RAVEN 1985, RAVEN & LUCAS 1985). To answer the above question, experiments were set up to study the effect of low and high NO<sub>3</sub><sup>-</sup>, in combination with low and high CO<sub>2</sub> during the growth period, on acidification,  $\Gamma_{CO_2}$  and R/P. It was hypothesized that high CO<sub>2</sub> and high NO<sub>3</sub><sup>-</sup> during growth would lead to a relatively high  $\Gamma_{CO_2}$  and a high R/P and the adverse conditions to low values for  $\Gamma_{CO_2}$  and R/P. The combination of low and high CO<sub>2</sub> and low and high NO<sub>3</sub><sup>-</sup> might also give insight in the mechanism, which induces acidification when CO<sub>2</sub> levels are low during growth. Under conditions of high NO<sub>3</sub><sup>-</sup>, low CO<sub>2</sub> will limit the rate of carbon fixation and thereby growth. Under conditions of low NO<sub>3</sub><sup>-</sup>, growth may be limited by N availability rather than by the rate of carbon fixation.

#### Materials and Methods

*Elodea canadensis* Michx. was grown in tanks on a clay substrate covered with sand (VAN GINKEL & PRINS 1998). For the present experiments sprouts of around 100 mm were transferred to 2-liter Erlenmeyer flasks containing an appropriate culture solution. Each flask contained about 15 sprouts, which were cultured for 1 to 3 weeks in 5% Hoagland growth medium. This solution contained 0.75 mM NO<sub>3</sub><sup>-</sup> (further referred to as high N) or 0.15 mM, one fifth of this concentration, (referred to as low N) at low or high CO<sub>2</sub>. Low CO<sub>2</sub> was obtained by continuous aeration with normal air and high CO<sub>2</sub> by constant titration of the growth medium to pH 6.6 by bubbling with CO<sub>2</sub> (ELZENGA & PRINS 1989). The medium was changed every fourth day. The temperature was 21 °C ( $\pm$  1 °C) and the fluence rate 60  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PAR with a photoperiod of 12 hours.

Rubisco and PEPCase were assayed in a crude extract made from newly formed shoots. The extraction buffer contained 60 mM HEPES, 24 mM MgCl<sub>2</sub>, 30 mM KHCO<sub>3</sub>, 0.24 mM NaEDTA and 6 mM DTT. The pH was set to pH 8.0 by KOH for Rubisco and to pH 7.1 for PEP-Case (adapted from REISKIND & BOWES 1991). The shoots were first frozen in liquid N<sub>2</sub> and thereafter ground in the presence of 5% (w/w) Polyclar-AT<sup>(TM)</sup>. The ground plant material was suspended in extraction buffer of pH 7.1 or 8.0. The ratio plant material/extraction buffer was 1:10 (w/v). Three aliquots of 50  $\mu\text{l}$  were taken and added to 950  $\mu\text{l}$  85% acetone for chlorophyll determination (LICHTENTHALER 1987). Thereafter the plant debris was spun down in an Eppendorf centrifuge (14,000 rpm, 10 min.). Aliquots of 100  $\mu\text{l}$  supernatant were tested for enzyme activity. The

assay for both enzymes was spectrophotometrically and based on a decrease in NADH concentration, measured as absorption at 340 nm. The Rubisco assay was as described by BESFORD 1983. The PEPCase assay was based on a method described by STITT & al. 1989. The buffer used for this assay contained 50 mM HEPES, 20 mM MgCl<sub>2</sub>, 25 mM KHCO<sub>3</sub>, 0.2 mM NaEDTA, 5 mM DTT and was brought to pH 8.0 with KOH. The reaction was started by adding phosphoenolpyruvate (PEP, final concentration 8 mM) after a preincubation at 30 °C. The enzyme activity was determined after 10 minutes incubation at 30 °C. The  $I_{CO_2}$  (at 21 % O<sub>2</sub>) was determined with an infra red gas analyzer (HOLADAY & al. 1983). The experimental solution was a 5% Hoagland medium buffered with 50 mM MES brought to pH 5.5 with KOH. This pH was chosen so that CO<sub>2</sub> was the only carbon source and to prevent any contribution by HCO<sub>3</sub><sup>-</sup> (VAN & al. 1976). During the experiment the pH remained constant, the fluence rate was 300 μmol m<sup>-2</sup> s<sup>-1</sup> PAR and the temperature 21 °C. The light induced pH-polar reaction was measured in artificial pond water, APW, containing 5 mM CaCl<sub>2</sub>, 1.5 mM KCl, 1 mM NaCl and KHCO<sub>3</sub> as indicated (PRINS & al. 1982).

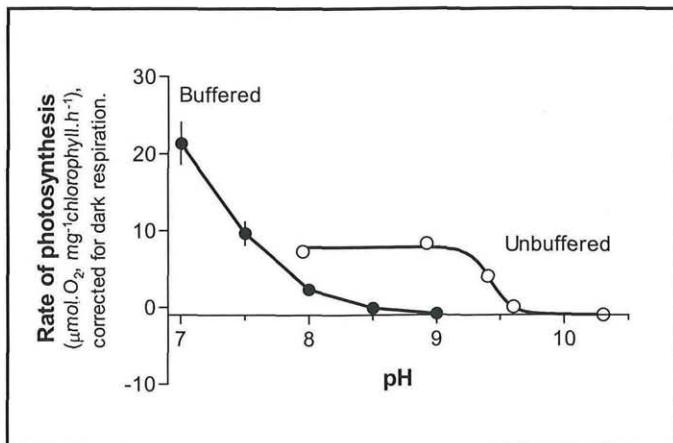


Fig. 1. Light induced O<sub>2</sub> production rate, corrected for dark respiration, of *E. canadensis* leaves plotted against pH using either a strongly buffered (25 mM MES/BTP) or an unbuffered APW solution containing 1 mM HCO<sub>3</sub><sup>-</sup>.

Photosynthetic O<sub>2</sub> production at different pH's was measured using an O<sub>2</sub> electrode. The experimental solution, consisting of APW and 25 mM MES/BTP buffer, was rapidly stirred. The combination of stirring and strong buffering should effectively prevent any use of HCO<sub>3</sub><sup>-</sup> as carbon source, as the build up of an acid region by polarity was prevented. To test this, net O<sub>2</sub> production was measured in a strongly buffered and an unbuffered experimental solution of APW. With buffering, O<sub>2</sub> production stopped completely at pH 8.5, while in APW it continued till pH 9.5 (Fig. 1).

## Results

The water in the tanks, wherein the plants originally were grown before the low and high CO<sub>2</sub> culture period, was weakly aerated by normal air and has to be considered 'low CO<sub>2</sub>'. Accordingly the plant material directly taken from the tanks showed pH-polarity, with abaxial acidification under CO<sub>2</sub> limiting conditions. Transferred to the Hoagland growth medium the *Elodea* sprouts grew vigorously,

especially in the high CO<sub>2</sub> solutions, and new sprouts (and roots) developed. The pH of the low CO<sub>2</sub> growth medium increased to values of around 10 in the light and returned in the dark to around 7, as shown in the inset of Fig. 2 for *Elodea* grown at low N. This drift of the pH, while the solution was aerated with normal air, indicated a high capacity of *Elodea* for extracting DIC from its surrounding medium (ADAMEC 1993, SAND-JENSEN & GORDON 1986). In accordance herewith *Elodea*, grown in 5% Hoagland under low CO<sub>2</sub> showed the typical pH polar reaction, when transferred to APW + 1 mM KHCO<sub>3</sub> pH 8, light 60 μmol m<sup>-2</sup> s<sup>-1</sup>, (Fig. 2) as described earlier (ELZENGA & PRINS 1989). When *Elodea* was grown under high instead of low CO<sub>2</sub> conditions no pH polarity was induced and there was only a small a-polar increase of the pH near both leaf sides and the change to adaxial alkalization and abaxial acidification did not occur (Fig. 2). At this stage no difference between low and high N plants was observed.

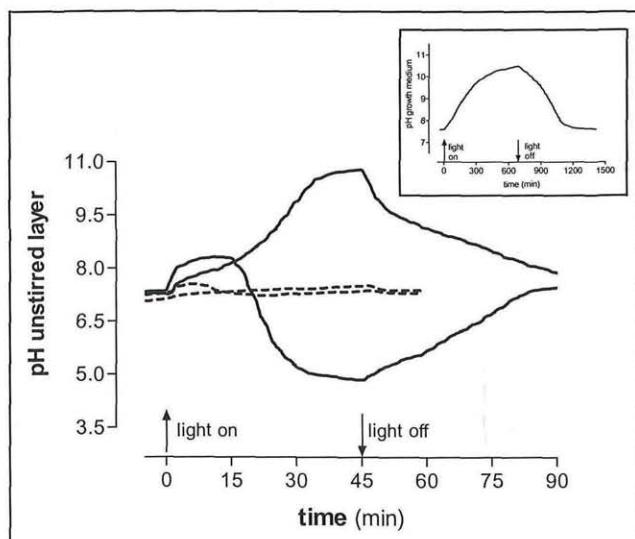


Fig. 2. pH-polar reaction of *E. canadensis* leaves grown at either low or high CO<sub>2</sub>. Medium: 5% Hoagland low N, fluence rate 60 μmol m<sup>-2</sup> s<sup>-1</sup> PAR. Inset: Culture solution pH of low CO<sub>2</sub> culture, one light-dark cycle (24 hours).

A shift to a C<sub>4</sub>-like state was observed in *Elodea* when the plants were cultured under low CO<sub>2</sub> conditions. The  $\gamma_{\text{CO}_2}$  was low for the low CO<sub>2</sub> plants compared to the  $\gamma_{\text{CO}_2}$  of plants cultured at high CO<sub>2</sub>. In the low CO<sub>2</sub> plants R/P shifted from Rubisco to PEPCase (Table 1). Fig. 3 shows the actual Rubisco and PEPCase activity for plants after being transferred to the Hoagland culture solutions of low and high N and C for 1 to 2 weeks. The results for high and low N are described below.

Table. 1.  $\Gamma_{\text{CO}_2}$  in ( $\mu\text{l l}^{-1} \pm \text{SEM}$ ), chlorophyll content in mg g FW $^{-1} \pm \text{SEM}$ , c<sub>a</sub>/c<sub>b</sub> and R/P of *E. canadensis* grown in 5% Hoagland, normal or 1% NO<sub>3</sub><sup>-</sup> (high or low N) for 2-3 weeks, under high and low CO<sub>2</sub> availability. \*) The  $\Gamma_{\text{CO}_2}$  for low CO<sub>2</sub> plants at either high or low N was not significantly different.

	High CO <sub>2</sub> High N	Low CO <sub>2</sub> High N	High CO <sub>2</sub> Low N	Low CO <sub>2</sub> Low N
Chlorophyll	2.53 ± 0.34	3.27 ± 0.36	2.31 ± 0.16	2.61 ± 0.18
Ca/Cb	2.77 ± 0.02	2.69 ± 0.05	2.79 ± 0.03	2.68 ± 0.02
$\Gamma_{\text{CO}_2}$	70-90	20-30*)	70-90	30-40*)
R/P	2.09 ± 0.35	1.25 ± 0.41	3.10 ± 0.33	1.88 ± 0.17

The Rubisco activity remained constant in the low CO<sub>2</sub>, but increased in the high CO<sub>2</sub> plants. The PEPCase activity in contrast, remained nearly constant in the high as well as in the low CO<sub>2</sub> plants (Fig. 3). As a result the R/P was relatively high in these high N plants grown at high CO<sub>2</sub>, but low in those grown at low CO<sub>2</sub> (Fig. 3; Table 1). This corresponded with a high  $\Gamma_{\text{CO}_2}$  in the high CO<sub>2</sub> plants and a low  $\Gamma_{\text{CO}_2}$  in the low CO<sub>2</sub> plants (Table 1).

The Rubisco activity increased in both low and high CO<sub>2</sub> grown plants (Fig. 3). The PEPCase activity increased in the low CO<sub>2</sub> plants, while it showed only a slight, probably not significant, downward tendency in the high CO<sub>2</sub> plants (Fig. 3). As a result the activity ratio of Rubisco/PEPCase (R/P) of the low N plants showed the same tendency as in high N plants, R/P was high in high CO<sub>2</sub> grown plants and low in those grown at low CO<sub>2</sub> (Fig. 3). As with the high N plants this corresponded with a high  $\Gamma_{\text{CO}_2}$  in the high CO<sub>2</sub> plants and a low  $\Gamma_{\text{CO}_2}$  in those grown at low CO<sub>2</sub> (Table 1).

Net O<sub>2</sub> production by Elodea was measured, using a strongly buffered experimental solution with a constant DIC concentration of 1 mM and a varying pH. Under these conditions photosynthesis depends entirely dependent on the CO<sub>2</sub> with no contribution of HCO<sub>3</sub><sup>-</sup> (Fig. 4). Under these conditions no difference was observed between the low and high CO<sub>2</sub> plants, of either low or high N grown plants. Therefore the low and high CO<sub>2</sub> data were pooled. In contrast there was a very marked difference between the low and high N plants. The rate of O<sub>2</sub> production of the high N plants at the highest external [CO<sub>2</sub>] was twice that of the low N plants. In the low N plants photosynthetic O<sub>2</sub> production was saturated between 0.1 and 0.2 mM CO<sub>2</sub> while in the high N plants saturation was not yet reached at 0.5 mM CO<sub>2</sub>.

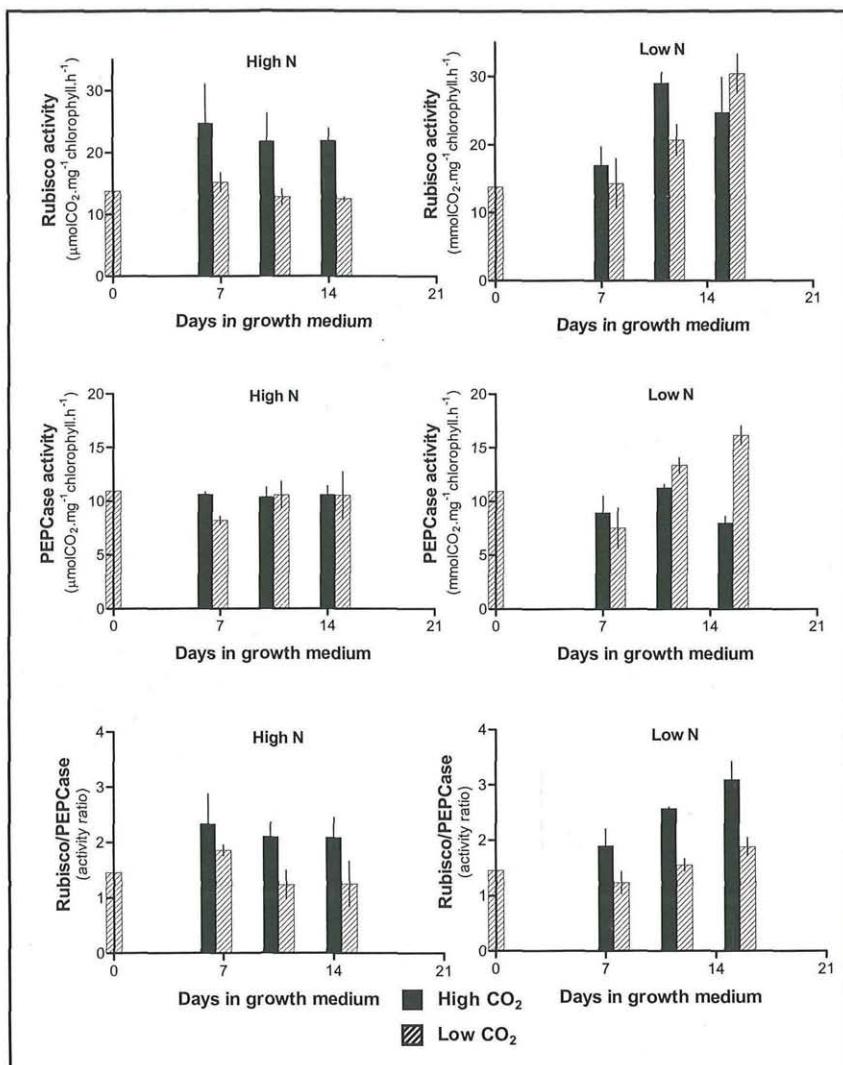


Fig. 3. Rubisco and PEPCase activity of *E. canadensis* shoots grown in 5% Hoagland solution for 1-2 weeks, at high or low N and low or high  $\text{CO}_2$ , fluence rate  $60 \mu\text{mol m}^{-2} \text{s}^{-1}$  PAR, photoperiod 12 h, 21 °C. 1-2 weeks.

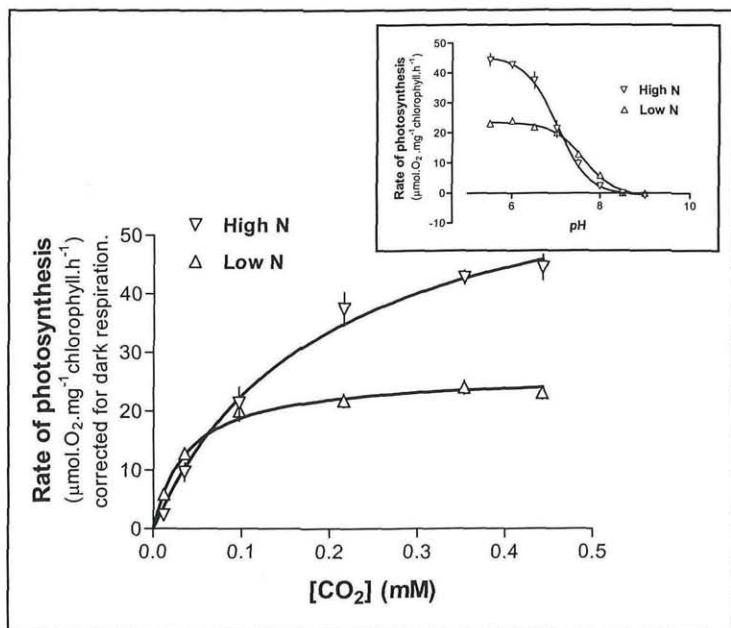


Fig. 4. Light induced  $O_2$  evolution of *E. canadensis* leaves at  $60 \mu\text{mol m}^{-2} \text{s}^{-1}$  versus  $[\text{CO}_2]$ ,  $[\text{DIC}]$  was constant,  $1 \text{ mM}$ .  $[\text{CO}_2]$  was varied by changing the pH, see inset.

The chlorophyll content and the chlorophyll a/b ratio,  $c_a/c_b$ , depended on the growth conditions ( $N$ ,  $\text{CO}_2$ ) of the plants. High  $\text{CO}_2$  plants grown at either low or high  $N$ , did not significantly differ in their chlorophyll content. When plants were grown at low  $\text{CO}_2$  high  $N$  plants contained far more chlorophyll than low  $N$  plants, and also more than the plants grown at high  $\text{CO}_2$ , either at low or high  $N$ . A higher  $c_a/c_b$  was observed in plants grown at high  $\text{CO}_2$  compared to plants grown at low  $\text{CO}_2$  (Table 1). The different growth conditions resulted in plants with similar dry/fresh weight ratios of around 8%. Furthermore both, low and high  $\text{CO}_2$  plants (leaves) had a similar protein content. The low  $N$  plants, both low and high  $\text{CO}_2$ , had a lower protein content,  $3-4 \mu\text{g protein g FW}^{-1}$ , than high  $N$  plants,  $7-8 \mu\text{g protein g FW}^{-1}$ .

## Discussion

Light induced pH polarity is suppressed in *Elodea* grown at high  $\text{CO}_2$  in both low (Fig. 2) and high  $N$  cultured plants (data not shown, but see ELZENGA & PRINS 1989), in agreement with the inhibition of bicarbonate utilization (ADEMEC 1993). At  $0.4 \text{ mM}$  external  $[\text{CO}_2]$  the rate of  $O_2$  production of high  $N$  grown plants was twice that of low  $N$  plants. No difference was observed between low and high

CO<sub>2</sub> plants. Strong pH buffering makes acidification by pH-polarity ineffective and therefore, CO<sub>2</sub> was the only C source in these experiments (PRINS & al. 1982). The stimulation of O<sub>2</sub> production by high N growth conditions did not result from an increased chlorophyll content as the rate of O<sub>2</sub> production is expressed on a chlorophyll basis.

High CO<sub>2</sub> growth conditions induced a high  $\Gamma_{\text{CO}_2}$ , in both high and low N plants (JAHNKE & al. 1991, ADAMEC 1993). The compensation point of low CO<sub>2</sub> grown *Elodea* seemed somewhat higher in high than in low N grown plants. This may be related to a higher rate of net O<sub>2</sub> production observed in high N grown plants and a twice as high protein content of these leaves.

The data on Rubisco and PEPCase activity show that R/P was higher in the high than in low CO<sub>2</sub> plants. The underlying changes in Rubisco and PEPCase activity differ between low and high N *Elodea* plants. The growth condition in the tanks, from which the original starting plant material was collected, was low CO<sub>2</sub>. Accordingly the Rubisco activity increased under high CO<sub>2</sub> in high N plants conditions while it remained the same in low CO<sub>2</sub> plants. The PEPCase activity of the high N plants did change however, neither in the low CO<sub>2</sub> nor in the high CO<sub>2</sub> plants. As a result R/P increased when plants were grown under high N, high CO<sub>2</sub> conditions. In the low N plants the changes are less clear although they lead to a similar change in R/P: Rubisco activity increased under both low and high CO<sub>2</sub> conditions; PEPCase activity of low N plants only increased in the low CO<sub>2</sub> grown plants. No significant change was observed in high CO<sub>2</sub> grown plants. The combined effect is an increase of R/P in the low N, high CO<sub>2</sub> grown plants. An observed increase of both Rubisco and PEPCase activity under low CO<sub>2</sub> conditions in low N plants indicates a more specific effect of low N during growth.

*E. canadensis* is taxonomically closely related to *H. verticillata*, both species belong to the Hydrocharitaceae family. The species are morphologically very similar, both are fresh water submerged macrophytes and both show photosynthetic HCO<sub>3</sub><sup>-</sup> utilization and pH polarity (ELZENGA & PRINS 1986, 1989, REISKIND & al. 1997). In *Hydrilla* C<sub>4</sub>-like characteristics were induced by so-called summer growth conditions: high light and temperature and low CO<sub>2</sub> (HOLADAY & al. 1983, BOWES & SALVUCCI 1989). Typical for C<sub>4</sub> like *Hydrilla* are a low  $\Gamma_{\text{CO}_2}$  and a low R/P activity ratio due to a relatively high PEPCase activity compared to C<sub>3</sub> like *Hydrilla*. In *Elodea* pH polarity and the C<sub>4</sub> characteristics of a low  $\Gamma_{\text{CO}_2}$  and a relatively high R/P ratio were all induced by growing the plants under low CO<sub>2</sub> conditions, while fluence rate and temperature were kept constant. This effect of high CO<sub>2</sub> growth conditions may seem comparable to some extent to the effect of winter and summer conditions on *Hydrilla*. One of the characteristics of winter conditions, which lead to C<sub>3</sub> plants characterized by a high  $\Gamma_{\text{CO}_2}$ , is a higher availability of CO<sub>2</sub> compared to summer conditions which lead to C<sub>4</sub> plants with a low  $\Gamma_{\text{CO}_2}$ , (SALVUCCI & BOWES 1981, HOLADAY & al. 1983, BOWES & SALVUCCI 1989). However, winter growth conditions did not suppress pH-polarity in *Hydrilla*. The observation of local acidification, leading to the conversion of HCO<sub>3</sub><sup>-</sup> into CO<sub>2</sub>, and C<sub>4</sub>-like characteristics induced in both species by low CO<sub>2</sub>, raises the question how these two processes cooperate or interact. Given the similarities between both spe-

cies it is assumed that the mechanism of CCM in *Hydrilla* is also active in *Elodea*. Activity of such a C<sub>4</sub>-like CCM could result in a lowering of cytoplasmic CO<sub>2</sub> producing a steeper gradient for CO<sub>2</sub> between apoplast and cells leading to a higher photosynthetic rate. Although this seems an attractive hypothesis it is not entirely supported by the experiments on photosynthetic O<sub>2</sub> production. The O<sub>2</sub> production as determined in these experiments reflected CO<sub>2</sub> fixation without HCO<sub>3</sub><sup>-</sup> utilization. Under conditions of an active CCM therefore, one would expect a higher rate of CO<sub>2</sub> fixation and thus a higher rate of O<sub>2</sub> production by low CO<sub>2</sub> plants. This was not observed, neither in the low N nor in the high N plants. Assuming that CCM also occurred in *Elodea* it did not result in a higher rate of O<sub>2</sub> production, in contrast to the use of HCO<sub>3</sub><sup>-</sup> via acidification, which leads to an increased O<sub>2</sub> production (PRINS & al. 1982). Clearly the two mechanisms, CCM and pH-polarity, contribute in different ways to photosynthesis in SAM like *Elodea* and *Hydrilla*. CCM activity seems to result in suppression of photorespiration but has only a marginal effect, if any, on the net rate of photosynthesis when diffusion of CO<sub>2</sub> into the leaf is limiting.

The cause of the stimulatory effect of high N during growth on photosynthetic O<sub>2</sub> production in both low and high CO<sub>2</sub> plants remains unclear. Possibly low N during growth in low CO<sub>2</sub>, but not in high CO<sub>2</sub>, induces a C<sub>4</sub>-like state, i.e. it lowers the  $\Gamma_{CO_2}$  and the R/P ratio, which may indicate that suppression of photorespiration is related to a more efficient use of N (RAVEN 1985). MADSEN & BAATTTRUP-PEDERSEN 1995 showed that in *Elodea* Rubisco activity was directly related to tissue-N level, and activity was substantially higher than could be expected on the basis of photosynthetic capacity. Furthermore, the relation between initial slope of the CO<sub>2</sub> response curve and Rubisco activity was linear. So clearly Rubisco had a regulatory effect under CO<sub>2</sub>-limiting circumstances and a similar conclusion can be drawn from the present results.

#### A c k n o w l e d g e m e n t

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